

Case Report
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BCRP Expression with Lipofuscin Accumulation in Abnormal Neurons from a Child with Transmantle Cortical Dysplasia (TMCD) and Refractory Epilepsy

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ABSTRACT

Cerebral cortical development's malformations, including the transmantle cortical dysplasia (TMCD), have been associated with refractory epilepsy (RE). Several ABC-transporters as "P-glycoprotein (P-gp), Multidrug resistance proteins (MRP-1) and breast Cancer Resistant Protein (BCRP)" are up-regulated in human epileptogenic brain lesions of RE, however they have not been explored in Transmantle cortical dysplasia (TMCD).

We describe a 13 years old boy with Refractory Epilepsy (RE) and abnormal Magnetic Resonance Image (MRI) (T1, FLAIR and T2) compatible with TMCD. Clinical follow-up, images and pathologic studies were developed by routinely methods. Epilepsy surgical treatment included total lesion resection with complete seizures remission at date.

Deeper brain areas related with images findings, showed features of TMCD with abnormal ballooned neurons with high accumulation of PAS+, sudanophilic and autofluorescent lipopigment (LP). Immunohistochemistry, using primary monoclonal antibodies for P-gp, MVP and BCRP proteins, showed high expression of BCRP in several ballooned-LP+ cells. In contrast, P-gp and MVP were negative and MRP-1 has not been investigated.

The links of BCRP with LP and AEDs are not known, however, the expression of BCRP in these LP+ ballooned neurons from the epileptogenic brain area, with P-gp/MVP negative results, suggest that BCRP could be associated to refractory epileptic phenotype.

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Introduction

Several malformations of cortical development have been described associated with refractory epilepsy and developmental delay. The main malformations identified were heterotopic gray matter, cortical tubers, focal cortical dysplasia, polymicrogyria, agyria-pachygyria, schizencephaly/cleft, transmantle dysplasia, and hemimegalencephaly [1, 2].

The transmantle cortical dysplasia (TMCD) was reported as a specific anomaly resulting from abnormal stem cell development, representing 5% of the main malformations identified [3,4]. TMCD was first described in 18 patients younger than 20 years with epilepsy or fixed neurologic deficits that presented MRI signal abnormalities extending from the cortex to the superolateral wall

of the lateral ventricle. The histological features of their brain biopsies are characterized by cortical disorganization, neuronal cytomegaly, balloon cells, indistinct cortical gray matter-white matter junctions, and variable accompanying astrogliosis. A specific anomaly resulting from abnormal stem cell development was proposed and [3].

Similar malformations of cortical development have been also described as additional lesions in tuberous sclerosis complex [5,6].

Two proteins associated with multidrug resistance in cancer, P-glycoprotein (P-gp) and multidrug resistance-associated protein 1 (encoded by ABCB1 and ABCC1 genes respectively), are up-regulated in human epileptogenic pathologies and the molecular basis of drug resistance in epilepsy is being intensively explored in experimental epilepsy models [7-21].

Another transporter named as “brain multidrug resistance protein” (BMDP) has been discovered at the porcine blood brain barrier (BBB) and was shown to be highly homologous to the human breast cancer resistance protein (BCRP), which is an other ABC transporter that confer multidrug resistance phenotype to the expressive cells [22-25]. However at date, all studies comparing BCRP expression in control and epileptic human brain tissue demonstrated the constitutive expression of BCRP in the brain capillary endothelium, but these data do not show differences in BCRP expression levels between the groups. Due to the current lack of evidence on BCRP overexpression in human epileptic brain tissue, BCRP is unlikely a major player in ASD resistance as proposed by the transporter hypothesis [26]. Here we describe at the first time the high BCRP expression in abnormal neurons loaded with lipofuscin from epileptogenic brain area. in a pediatric case of refractory epilepsy due TMCD.

Case Report

We describe a 13 years old boy with right focal clonic seizures (upper members and half body) from 5 month of life, without others personal or familial antecedents. Initial neurological examination, laboratory studies, EEG and CT scans were normal. Isolated crisis persisted thorough 8 years with normal IQ, normal intercritic EEG or left focal spikes. After this age, the number and intensity of seizures were increased without control with different antiepileptic drug schedules, and an other CT scan and MRI without abnormalities. (not shown)

Actually, at 13 years old, he present a chronic story of daily multiple crisis, light left hemiparesia and IQ = 75.

A recent MRI study indicated the engrossment of left frontal cortex, associated to an brain area with light signals changes related to gray matter, extended to subjacent left ventricle (hyper-intense in T1 and I/R, hypo-intense in FLAIR and iso-intense in T2), compatible with diagnosis of TMCD (Figure 1).



Figure 1: RMN -T2 showing the epileptogenic TMCD

The surgical resection of the brain epileptogenic lesion areas were developed and now, the patient remains without crisis since one year ago.

The brain material was examined by routinely histochemical methods, electronic microscopy examination, and used to investigate the expression of P-gp, MVP-1 and BCRP transporters.

Methods

Brain Tissue Samples

Surgical specimens of brain tissues were selected from the tissue collection of the Pathology Laboratory of the Garrahan Children's

Hospital of Buenos Aires. Patient had been surgically treated for intractable epilepsy, and brain samples were surplus to diagnostic requirements.

Morphological Analysis

Brain tissue was fixed in 10% buffered formalin and embedded in paraffin. Sections were stained with hematoxylin-eosin, Nissl, Luxol-Fast-Blue with PAS and Sudan Black methods, for morphological analysis.

Immunohistochemistry Methods

• Drug Transporters: The monoclonal antibodies and dilutions were used as follows: P-gp (1:50, clone JSB-1; Novocastra, Newcastle Upon Tyne, UK), MVP (1:50, Signet-Dedham, MA), and BCRP (1:50, Kamiya Biomed.Co.- Seattle). MRP-1 was not investigated.

• Secondary polyclonal antibody was performed with a Streptavidin immunoperoxidase kit, according to the protocol recommended by the manufacturer (Biogenix, San Ramón, CA, USA).

Results

A-Pathological Findings

Biopsy evidenced normal left frontal cortex and a deep abnormal area with totally ballooned neurons loaded with PAS+ sudanophilic and autofluorescent compound, corresponding to lipopigment likes to neuronal ceroid lipofiscinosis. (Figure 2a)

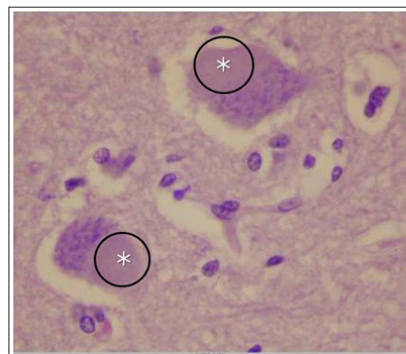


Figure 2: Hematoxylin-Eosine staining: Ballooned neurons with lipofuscin-like lipopigment accumulation (*)

Electronic Microscopy

Granular-dense and vacuolar electron-clear material with characteristics of classic lipofuscine are observed (Figure 3 Magnifications: a- 4000X, b-22000X)

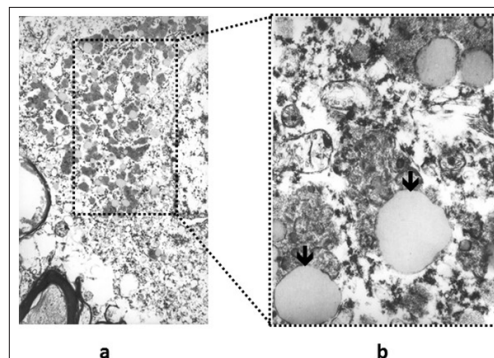


Figure 3: vacuoles are indicated by a red arrow (b)

B-Immunohistochemistry

Drug Transporters

P-glycoprotein (P-gp) was located in the luminal membrane of brain capillary endothelial cells without immunostaing differential

pattern compared with normal brain areas. Similarly, BCRP was highly expressed in BBB from both normal and pathological areas.

However, in brain parenchyma cells from epileptogenic lesion, P-gp and MVP were not detected, but strikingly BCRP was highly expressed in the abnormal ballooned neurons, with a particular polarized distribution in opposite side of lipopigment accumulation (Figure 4).

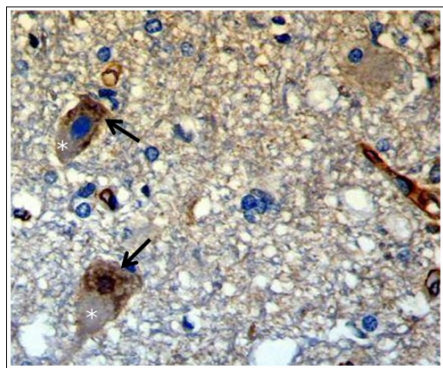


Figure 4: BCRP IHC: Ballooned neurons presenting polarized negative areas with lipid accumulation (*) and positive immunostaining for BCRP in the opposite site (→)

The lipofuscin-like lipopigment accumulation was restricted to abnormal brain area of the mantle dysplasia without diffuse distribution as observed in neuronal ceroid lipofuscinosis or physiologic aging, and unusually it was accumulated in abnormal neurons.

Discussion

The peculiar cytological features of the ballooned abnormal neurons observed in the specimens from the epileptogenic brain lesion of our patient, are not compatible with previously described in ballooned cells from Taylor's cortical dysplasia, tuberous sclerosis, or the findings observed in aging and it was not according with previously described features of childhood neuronal ceroid-lipofuscinosis in Argentina [27]. Intensive lipopigment accumulation was restricted to the abnormal cells from the lesion's area, without diffuse and/or extensive distributive pattern as described in others conditions with ceroid lipofuscin accumulation [28,30].

The clinical and brain images features, correlated with a cortical developmental disease compatible with a transmantle cortical dysplasia and refractory epilepsy [3,4].

The seizures abrogates with the surgical treatment by resection of the abnormal brain area, indicates that the particular morphological features correlated with the epileptogenic activity of the lesion.

At our knowledge, the blood-brain barrier (BBB) plays the predominant role in controlling the passage of endogenous and xenobiotic substances between the circulating blood and the extracellular fluid environment of the brain. So far, the multidrug resistance in epilepsy has been almost exclusively attributed to MDR-1 gene encoded P-glycoprotein (P-gp), the most prominent member of the ATP-binding cassette (ABC) transporter family, linked to higher expression in the luminal membrane of brain capillary endothelial cells and also expressed in brain parenchyma cells including neurons and astrocytes [7-21].

More recently, a called "brain multidrug resistance protein" (BMDP) has been discovered at the porcine BBB. Phylogenetic

analysis and multiple sequence alignment showed that porcine BMDP is most related to the human and mouse breast cancer resistance protein (BCRP) [22,31].

It was demonstrated by Immunofluorescence confocal microscopy that BCRP is normally located at the blood-brain barrier, mainly at the luminal surface of microvessel endothelium resembling that of P-gp. Because both transporters have several common substrates, BCRP may give an additional barrier to drug access to the brain [32,33]. However, at date the relationship between BCRP and AEDs still remains to be clarified.

It was reported that BCRP is expressed ubiquitously in brain capillary endothelium in patients with RE, but the authors concluded that there was no qualitative up-regulation of this transporter [34].

More recently, Vogelgesang S et al have been demonstrated that BCRP was highly expressed in vascular endothelial cells (VEC) of BBB as well as in astrocytes from brain specimens of patients with dysembryoplastic neuroepithelial tumors and RE [35].

In our patient, the P-gp and MVP were negative in brain parenchyma cells, and we can't study MRP-1 expression, however BCRP was strongly immunoreactive not only in VEC of BBB, but also in several abnormal neurons with high lipid accumulation, being the first observation of BCRP expressed in LP+ abnormal neurons associated with both epileptogenic and refractory phenotypes.

The relationship between the ABC transporter BCRP and LP has not been studied. In this regard, it important to notice that mutations on ABCA4 gene (an other member of ABC transporter family, also known as ABCR) has been demonstrated related with a Rod photoreceptor retinoid transport alteration and over 300 mutations in this ABCR gene have been associated with a variety of clinically distinct autosomal recessive retinal degenerative diseases, including Stargardt macular dystrophy, fundus flavimaculatus, cone-rod dystrophy, and retinitis pigmentosa, characterized by lipofuscin accumulation [36-40]. The lipofuscin accumulation is a features of ABCR gene mutation in Stargardt and also in the Age-Related Macular Degeneration diseases and it is coincident with the descriptions observed in the ABCR knockout mice, suggesting that similar mechanisms could be present in our case of the TMCD [41].

Our results suggest that BCRP expression in ballooned neurons can't protects this abnormal cells against the lipofuscine accumulation. We don't know if our patient have any ABCR mutation, or if he have any BCRP polymorphism associated with this dysfunctional lipid transport. However, it has been suggested that humans or animals with low or absent BCRP activity may be at increased risk for developing protoporphyria and diet-dependent phototoxicity. Interestingly, lipofuscine observed in our case was a fluorescent pigment, and the primary pathologic defect in Stargardt's disease is accumulation of "toxic lipofuscin pigments" such as N-retinylidene-N-retinylethano-lamine (A2E) in cells of the retinal pigment epithelium. This accumulation appears to be responsible for the photoreceptor death and severe visual loss in Stargardt's patients. Recently it has probed that treatment with isotretinoin may inhibit LP accumulation and thus delay the onset of visual loss in Stargardt's patients, perhaps giving an alternative treatment in TMCD cases with lipofuscin accumulation as described here [42].

In summary, our case of TMCD showed a particular pattern of immunohistochemistry on brain parenchyma cells, characterized

by non-detectable P-gp/MVP proteins, but high BCRP expression in LP+ abnormal neurons from epileptogenic brain area.

These results suggest that BCRP could play a role in the development of refractory epilepsy phenotype.

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References

1. Crino PB (2004) Malformations of cortical development: molecular pathogenesis and experimental strategies. *Adv Exp Med Biol* 548: 175-191.
2. Kirchhof K, Harting I, Bast T, Seitz A (2003) Focal cortical dysplasias: neuroradiological findings and differential diagnosis. *Rofo Fortschr Geb Rontgenstr Neuen Bildgeb Verfahr* 175: 1056-1063.
3. Barkovich AJ, Kuzniecky RI, Bollen AW, Grant PE (1997) Focal transmantle dysplasia: a specific malformation of cortical development. *Neurology* 49: 1148-1152.
4. Leventer RJ, Phelan EM, Coleman LT, Kean MJ, Jackson GD, et al. (1999) Neurology. Clinical and imaging features of cortical malformations in childhood. *Neurology* 53: 715-722.
5. Christophe C, Sekhara T, Rypens F, Ziereisen F, Christiaens F, et al. (2000) MRI spectrum of cortical malformations in tuberous sclerosis complex. *Brain Dev* 22: 487-493.
6. Vigliano P, Canavese C, Bobba B, Genitori L, Papalia F, et al. (2002) Transmantle dysplasia in tuberous sclerosis: clinical features and surgical outcome in four children. *J Child Neurol* 17: 752-758.
7. Lazarowski A, Sevlever G, Taratuto A, Massaro M, Rabinowicz A (1999) Tuberous Sclerosis associated with MDR1 gene expression and drug-resistant epilepsy. *Pediatr Neurol* 21: 731-734.
8. Sisodiya SM, Lin WR, Harding BN, Squier MV, Thom M (2002) Drug resistance in epilepsy: expression of drug resistance proteins in common cause of refractory epilepsy. *Brain* 125: 22-31.
9. Tishler DM, Weinberg KI, Hinton DR, Barbaro N, Annett GM, et al. (1995) MDR1 gene expression in brain of patients with medically intractable epilepsy. *Epilepsia* 36: 1-6.
10. Dombrowski S, Desai S, Marroni M, L Cucullo, K Goodrich, et al. (2001) Overexpression of multiple drug resistance genes in endothelial cells from patients with refractory epilepsy. *Epilepsia* 42: 1501-1506.
11. Lazarowski A, Lubieniecki F, Camarero S, Pomata H, Bartuluchi M, et al. (2004) Multidrug resistance proteins in tuberous sclerosis and refractory epilepsy. *Pediatr Neurol* 30: 102-106.
12. Lazarowski A, Massaro M, Schteinschnaider A, Intruvini S, Sevlever G, et al. (2004) Neuronal MDR-1 gene expression and persistent low levels of anticonvulsants in a child with refractory epilepsy. *Ther Drug Monit* 26: 44-46.
13. Marroni M, Agrawal M, Kight K, Hallene KL, Hossain M, et al. (2003) Relationship between expression of multiple drug resistance proteins and p53 tumor suppressor gene proteins in human brain astrocytes. *Neuroscience* 121: 605-617.
14. Sisodiya SM (2003) Mechanisms of antiepileptic drug resistance. *Curr Opin Neurol* 16: 197-201.
15. Aronica E, Gorter JA, Jansen GH, van Veelen CW, van Rijen PC (2003) Expression and cellular distribution of multidrug transporter proteins in two major causes of medically intractable epilepsy: focal cortical dysplasia and glioneuronal tumors. *Neuroscience* 118: 417-429.
16. Potschka H, Fedrowitz M, Loscher W (2003) Multidrug resistance protein MRP2 contributes to blood-brain barrier function and restricts antiepileptic drug activity. *J Pharmacol Exp Ther* 306: 124-131.
17. Sills GJ, Kwan P, Butler E, de Lange EC, van den Berg DJ, et al. (2002) P-glycoprotein-mediated efflux of antiepileptic drugs: preliminary studies in *mdr1a* knockout mice. *Epilepsy Behav* 3: 427-432.
18. Rizzi M, Caccia S, Guiso G, Richichi C, Gorter JA et al. (2002) Limbic seizures induce P-glycoprotein in rodent brain: functional implications for pharmacoresistance. *Neurosci* 22: 5833-5839.
19. Volk HA, Burkhardt K, Potschka H, Chen J, Becker A, et al. (2004) Neuronal expression of the drug efflux transporter P-glycoprotein in the rat hippocampus after limbic seizures. *Neuroscience* 123: 751-759.
20. Lazarowski A, Girardi E, Ramos AJ, García-Rivelo H, Brusco A (2004) MDR-1 gene expression (Glycoprotein P-170) in different brain areas in an experimental epilepsy model. *J Epilepsy Clin Neurophysiol* 8: 101-104.
21. Lazarowski A, Ramos AJ, García-Rivello H, Brusco A., Girardi E (2004) Neuronal and glial expression of the multidrug resistance gene product in an experimental epilepsy model. *Cell Mol Neurobiol* 24: 77-85.
22. Eisenblatter T, Galla HJ (2002) A new multidrug resistance protein at the blood-brain barrier. *Biochem Biophys Res Commun* 293: 1273-1283.
23. Burger H, Van Tol H, Boersma AW, Brok M, Wiemer EA, et al. (2004) Imatinib mesylate (STI571) is a substrate for the breast cancer resistance protein (BCRP) / ABCG2 drug pump *Blood* 104: 2940-2942.
24. Suvannasankha A, Minderman H, O'Loughlin KL, Nakanishi T, Greco WR, et al. (2004) Breast cancer resistance protein (BCRP/MXR/ABCG2) in acute myeloid leukemia: discordance between expression and function. *Leukemia* 18: 1252-1257.
25. Sarkadi B, Ozvegy-Laczka C, Nemet K, Varadi A (2004) ABCG2 -a transporter for all seasons. *FEBS Lett* 567: 116-120.
26. Taratuto AL, Saccoliti M, Sevlever G, Ruggieri V, Arroyo H, et al. (1995) Childhood neuronal ceroid-lipofuscinoses in Argentina. *Am J Med Genet* 57: 144-149.
27. Tang F, Hartz AMS, Bauer B (2017) Drug-Resistant Epilepsy: Multiple Hypotheses, Few Answers. *Front Neurol* <https://doi.org/10.3389/fneur.2017.00301>
28. Goebel HH (1997) Morphologic diagnosis in neuronal ceroid lipofuscinosis. *Neuropediatrics* 28: 67-69.
29. Dyken P (1988) Reconsideration of the classification of the neuronal ceroid-lipofuscinoses. *Am J Med Genet* 5: 69-84.
30. Dyken P, Wisniewski K (1995) Classification of the neuronal ceroid-lipofuscinoses: an expansion of the atypical forms. *Am J Med Genet* 57: 150-154.
31. Eisenblatter T, Huwel S, Galla HJ (2003) Characterisation of the brain multidrug resistance protein (BMDP/ABCG2/BCRP) expressed at the blood-brain barrier. *Brain Res* 971: 221-231.
32. Zhang W, Mojsilovic-Petrovic J, Andrade MF, Zhang H, Ball M, et al. (2003) The expression and functional characterization of ABCG2 in brain endothelial cells and vessels. *FASEB J* 17: 2085-2087.
33. Cooray HC, Blackmore CG, Maskell L, Barrand MA (2002) Localisation of breast cancer resistance protein in microvessel endothelium of human brain. *Neuroreport* 13: 2059-2063.
34. Sisodiya SM, Martinian L, Scheffer GL, Van der Valk P,

- Cross JH, et al. (2003) Major vault protein, a marker of drug resistance, is upregulated in refractory epilepsy. *Epilepsia* 44:1388-1396.
35. Vogelgesang S, Kunert-Keil C, Cascorbi I, Mosyagin I, Schroder E, et al. (2004). Expression of multidrug transporters in dysembryoplastic neuroepithelial tumors causing intractable epilepsy. *Clin Neuropathol* 23: 223-231.
36. Allikmets R, Singh N, Sun H, Shroyer NF, Hutchinson A, et al. (1997) A photoreceptor cell-specific ATP-binding transporter gene (ABCR) is mutated in recessive Stargardt macular dystrophy. *Nat Genet* 15: 236-246.
37. Nasonkin I, Illing M, Koehler MR, Schmid M, Molday RS, et al. (1998) Mapping of the rod photoreceptor ABC transporter (ABCR) to 1p21-p22.1 and identification of novel mutations in Stargardt's disease. *Hum. Genet* 102: 21-26.
38. Martinez-Mir A, Paloma E, Allikmets R, Ayuso C, del Rio T, et al. (1998) Retinitis pigmentosa caused by a homozygous mutation in the Stargardt disease gene ABCR. *Nat Genet* 18: 11-12 .
39. Cremers FP, Van de Pol DJ, Van Driel M, Den Hollander A I, Van Haren FJ, et al. (1998) Autosomal recessive retinitis pigmentosa and cone-rod dystrophy caused by splice site mutations in the Stargardt's disease gene ABCR *Hum. Mol Genet* 7: 355-362.
40. Lewis R A, Shroyer N F, Singh N, Allikmets R, Hutchinson A, et al. (1999) Genotype/Phenotype analysis of a photoreceptor-specific ATP-binding cassette transporter gene, ABCR, in Stargardt disease. *Am. J Hum Genet* 64: 422-434.
41. Mata NL, Tzekov RT, Liu X, Weng J, Birch DG, et al. (2001) Delayed dark-adaptation and lipofuscin accumulation in *abcr*^{+/-} mice: implications for involvement of ABCR in age-related macular degeneration. *Invest Ophthalmol Vis Sci* 42: 1685-1690.
42. Radu RA, Mata NL, Nusinowitz S, Liu X, Sieving PA, et al. (2003) Treatment with isotretinoin inhibits lipofuscin accumulation in a mouse model of recessive Stargardt's macular degeneration. *PNAS* 100: 4742-4747.